Molecular Evolution Answering question 2

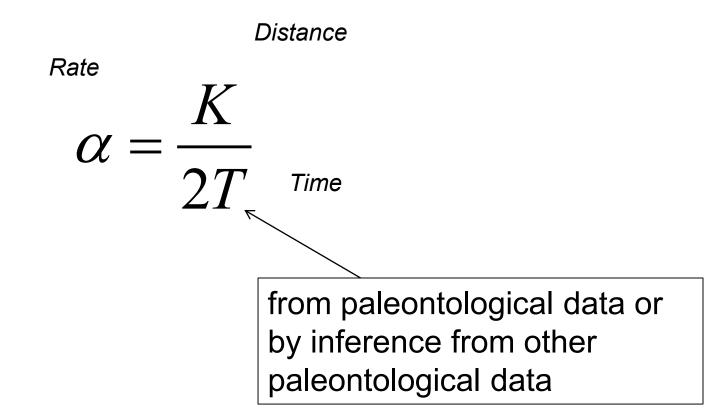
The degree of divergence between 2 sequences is the Hamming distance (the edit distance /length of the sequence)

Variables

- Number of mutations (K)
- Rate of mutation (α)
- Time elapsed since divergence (T)

Remember from Junior High Algebra: time x rate = distance

When divergence is neither too recent nor too remote in time*:



*Polymorphism prior to divergence in very close species Increased probability of same site multiple substitutions in remote species

Divergent Sequences Number of Mutations (Evolutionary Distance)

- We really don't know how many mutations have occurred in divergent sequences
 - There can be additional mutations of the same site in one sequence
 - The same site can mutate in both sequences
 - The same site in both sequences can mutate to the same base and appear never to have diverged

Divergent sequences-

Some possible mutation schemes*

Divergent sequence 1	original	Divergent sequence 2
·	A	
	С	
Т	Τ	$\rightarrow A$
$T \leftarrow C$	$\leftarrow A$	A
С	С	С
$C \leftarrow$	<i>G</i>	$\rightarrow T$
$T \leftarrow$	A	$\longrightarrow T$
Т	Т	Т
$G \leftarrow A$	← T —	$\rightarrow G$
G	G	G
A	$A \rightarrow$	$T \rightarrow A$
	A	
	С	
	Т	
	G	

*from Grauer and Li

Jukes and Cantor Mutation Model

- If a sequence exists over *t*, the probability of the base, say, A, at any given site being the same is pAA_t
- The joint probability that two (divergent) sequences having the same base at the same site is $p(A_0A_t)$ for seq1 × $p(A_0A_t)$ for seq 1, or $p^2A_0A_t$
- Likewise, the probability that two (divergent) sequences having a *different* base at the same site is p²AC_t or p²AG_t or p²AT_t
- The total probability is

$$p_{total} = p^2 A_0 A_t + p^2 A_0 C_t + p^2 A_0 G_t + p^2 A_0 T_t$$

Recalling that, for the Jukes and Cantor model,

$$pA_0A_t = \frac{1}{4} + \left(\frac{3}{4}\right)e^{-4\alpha t}$$

And, having just established that

$$p_{total} = p^2 A_0 A_t + p^2 A_0 C_t + p^2 A_0 G_t + p^2 A_0 T_t$$

we determine that

$$p_{total} = \frac{1}{4} + \left(\frac{3}{4}\right) (e^{-4\alpha t})^2 = \frac{1}{4} + \frac{3}{4} e^{-8\alpha t}$$

Now, p_{total} is the probability that we end up with the <u>same</u> <u>nucleotide</u> as we started with, after *t*. For our investigation of divergent sequences, we are really looking for the probability that the nucleotide in a given site would be <u>different</u> after *t*.

That probability is, of course $p_{different}=1-p_{total}$, or

$$p_{different} = \frac{3}{4} (1 - e^{-8\alpha t})$$

By rewriting

 $p_{different} = \frac{3}{4} (1 - e^{-8\alpha t})$

we get

$$-8\alpha t = -\ln(1 - \frac{4}{3}p_{different})$$

But we cannot estimate α . We do know, however, that $3\alpha t$ is the rate of substitutions per site .

Let K represent the number of substitutions per site since the sequences diverged. For the Jukes-Cantor model,

Arbitrarily, set
$$K = 2(3\alpha t) = 6\alpha t$$
 or $K = -\frac{4}{3}6\alpha t$

Substituting K into the expression $-8\alpha t = -\ln(1 - \frac{4}{3}p)$ we get

$$-\frac{4}{3}K = -\ln(1 - \frac{4}{3}p)$$
$$K = -\frac{3}{4}\ln(1 - \frac{4}{3}p)$$

Estimating Evolutionary Distance

K is a proxy evolutionary distance. In the final analysis, α will need to be calibrated, most likely by biological observation

$$dist \approx -\frac{3}{4}\ln(1-\frac{4}{3}p)$$

where $p = fraction \ of \ changed \ nucleotides$
$$or \ p = \left(\frac{\# \ of \ changes}{length \ of \ sequence}\right)$$

Hamming distance is sometimes defined as the number of changes (same as edit distance) and sometimes as the number of changes/sequence length. Here p is the Hamming distance

EXAMPLE: Consider these two sequences

The edit distance is 2.

p is
$$2/8 = .25$$

Dist= -0.75ln[1-4/3(0.25)]
= 0.30035

When diverging sequences are far apart, distance K becomes unreliable because of sites involved more than once

Substitution rates

- Coding DNA
 - Synonymous substitutions: same AA
 - Nonsynonymous substitutions: different AA
- Non coding DNA
 - Data from UTRs, else scant data

Protein Coding

Synonymous and Nonsynonymous substitutions

- A #1 or #2 position can influence whether #3 will make a synonymous substitution
- Transitions are more frequently synonymous than transversions

All of which make the models significantly more complicated

Codons

- 4-fold degeneracy: any nucleotide in the 3rd position specifies the same AA
 - gly: GGA,GGC,GGG,GGU
- 2-fold degeneracy: two nucleotides in the 3rd position specifiy the same AA
 - glutamic acid: GAA,GAG
 - Only transversions are nonsynonymous
- Special case: 3 nucleotides code for the same AA
 ileu: AUA,AUC,AUU
- 3 AAs (ser,leu,arg) have 6 codons
- 2AAs (met (AUG) and try (UGG) have only 1 codon

Type of substitution *vis à vis* rate of substitution* (in substitutions/billion yrs)

	Non degenerate	Twofold degenerate	Fourfold degenerate
Transition	0.40	1.86	2.24
Transversion	0.38	0.38	1.47

*Table from Grauer and Li

Rates

Coding DNA

Non-synonymous α 0 substitutions /site /year

 γ interferon 3.1x10⁻⁹ substitutions /site /year

Synonymous up to 25 x higher rate

Substitution rates within genes

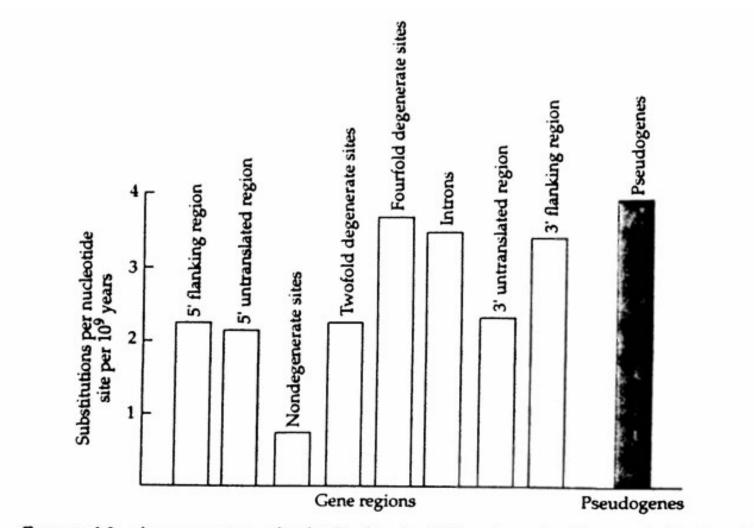


FIGURE 4.3 Average rates of substitution in different parts of genes (white) and i pseudogenes (gray). From Li (1997).

Mutation Rates

Possibly explained by

- Mutational input
- Genetic drift of neutral alleles
- Purifying selection against deleterious alleles (selectional constraint)

But what about positive selection?

If Darwinian positive selection, then K_{nonsynonymous} >K _{synonymous}

BUT

Statistical analysis does not lead to that conclusion

MOLECULAR CLOCK CONCEPT*

The assumption: Mutations occur at a fixed rate (α) across time

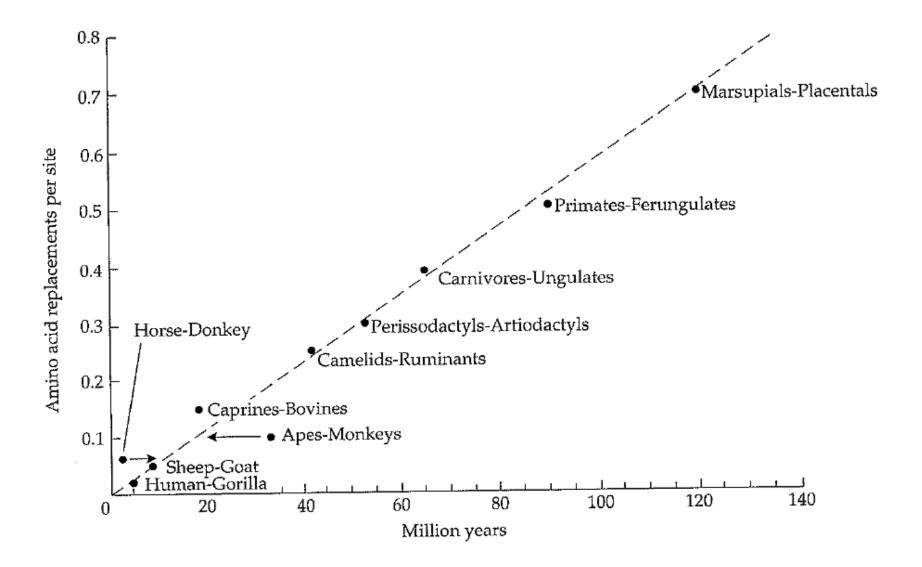
A theory, unproven. But, if indeed there is a molecular clock, then our formula

$$\alpha = \frac{K}{2T}$$

can be used when K is known but there are no paleontological data for T

*Important in phylogeny determinations

Molecular Clock in Action



Taken from Grauer and Li, modified from Langley and Fitch, 1974 Mol Evol 3 161-177 [4]